

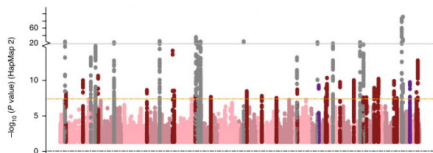
# Large-scale genome-wide enrichment analyses of 31 human complex traits

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University of Chicago

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# Examining associations between variables is a useful tool in genetics.

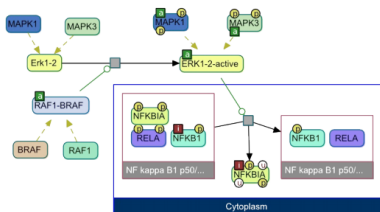
## GWAS: SNP~Phenotype



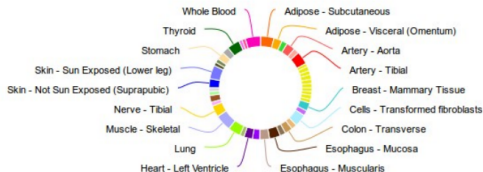
## 1000 Genomes: SNP~SNP



## Biological Pathways: Gene~Gene



## GTEX: Gene~Tissue



# Enrichment analysis combines multiple sources of association.

- **SNP-Trait:** GWAS summary statistics
- **SNP-SNP:** linkage disequilibrium (LD)
- **Gene-Gene:** biological pathways
- **Gene-Tissue:** RNA-seq across tissues

**Let's keep this talk "jargon-free".**

1. GWAS summary statistics
2. gene set enrichment analysis

# What are GWAS summary statistics?

- **Data:** phenotype  $Y$  and genotype  $X$
- **Size:**  $n$  (>10K) individuals and  $p$  (>1M) variants (SNPs)

$$Y := \begin{bmatrix} y_1 \\ y_2 \\ \vdots \\ y_n \end{bmatrix} \quad X := \begin{bmatrix} X_{11} & \dots & X_{1j} & \dots & X_{1p} \\ X_{21} & \dots & X_{2j} & \dots & X_{2p} \\ \vdots & \vdots & \vdots & \dots & \vdots \\ X_{n1} & \dots & X_{nj} & \dots & X_{np} \end{bmatrix}$$

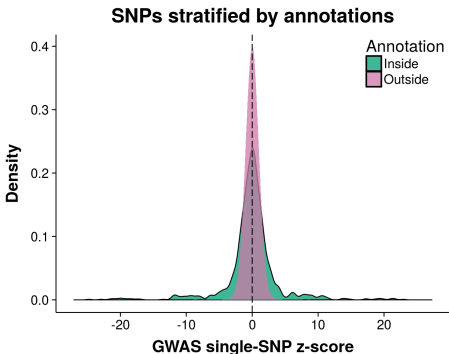
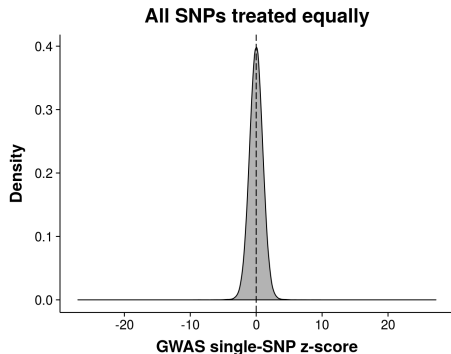
- **Model:** single-SNP association analysis

$$\begin{bmatrix} y_1 \\ y_2 \\ \vdots \\ y_n \end{bmatrix} \sim \begin{bmatrix} X_{1j} \\ X_{2j} \\ \vdots \\ X_{nj} \end{bmatrix} \rightsquigarrow \begin{cases} \hat{\beta}_j: & \text{marginal effect estimate} \\ \hat{\sigma}_j: & \text{standard error of } \hat{\beta}_j \end{cases}$$

- **Availability:**  $\{\hat{\beta}_j, \hat{\sigma}_j\} \gg \{Y, X\}$  (Nat. Genet. Editorial, 2012)

# What is enrichment analysis?

- **Phenotype:** low-density lipoprotein (Teslovich *et al.*, 2010)
- **Pathway:** *chylomicron-mediated lipid transport* (17 genes)
- **Annotation:** Is the SNP “near” a pathway gene? (yes or no)



**Recent reviews:** de Leeuw *et al.* (2016); Pers (2016); Mooney *et al.* (2014); Wang *et al.* (2010).

# The “enrichment” idea is simple, but there are (at least) two technical issues.

1. If the gene set is truly enriched, we should relax significance threshold for “green” SNPs, but how much to relax?
  - **Data-driven** threshold ← Function (Pathway, Phenotype)
  - Maintained type 1 error + Improved power
2. The “inflated” green curve may be driven by correlation between SNPs (LD), rather than enrichment of signal.
  - SNP 1 has a large genetic effect on a trait.
  - SNPs 2-100 have zero effect, but all are in **high LD** with SNP 1.
  - Thus, SNPs 1-100 all show very large **single-SNP** z-scores.

# We develop a method that systematically utilizes enrichment information.

Public Data

Inference

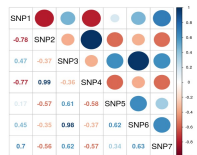
## GWAS summary statistics



$$\hat{\beta} := (\hat{\beta}_1, \dots, \hat{\beta}_p)^T, \quad \hat{S} := \text{diag}\{(\hat{s}_1, \dots, \hat{s}_p)^T\}$$

$\hat{\beta}_j$ : marginal effect estimate of SNP  $j$   
 $\hat{s}_j$ : standard error of  $\hat{\beta}_j$

## External LD estimates



$$\tilde{R} := [\hat{r}_{ij}]_{p \times p}, \quad \hat{r}_{ij}: \text{LD between SNP } i \text{ and } j$$

## Predefined gene sets

Gene	Chr	Start	End
BRAF	7	140419127	140624564
MAPK1	22	22108789	22221970
MAPK3	16	30125426	30134827
NFKB1	4	103422486	103538459

$$a_j := 1 \{ \text{SNP } j \text{ is "near" a gene in the set} \}$$

## Bayesian Model

### Likelihood function

$$L(\beta) := \text{Normal}(\hat{\beta}; \hat{S} \hat{R} \hat{S}^{-1} \beta, \hat{S} \hat{R} \hat{S})$$

### Prior distribution

$$\beta_j \sim \pi_j \cdot \text{Normal}(0, \sigma_\beta^2) + (1 - \pi_j) \delta_0$$

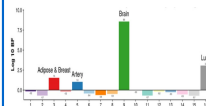
### Baseline model ( $M_0$ )

$$\log_{10} \left( \frac{\pi_j}{1 - \pi_j} \right) = \theta_0$$

### Enrichment model ( $M_1$ )

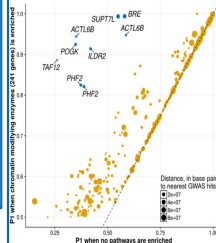
$$\log_{10} \left( \frac{\pi_j}{1 - \pi_j} \right) = \theta_0 + \alpha_j \theta$$

## Gene set enrichment



$$\text{BF}(\text{gene set}) = \frac{\Pr(\text{Data} | M_1)}{\Pr(\text{Data} | M_0)}$$

## Gene prioritization



$$\rho(\beta | \text{Data}, M_1) \text{ vs } \rho(\beta | \text{Data}, M_0)$$

# Address Issue 1: Learning enrichment from data

## Model-based approach:

- Assume that SNP  $j$  is trait-associated with probability  $\pi_j$
- Represent  $\pi_j$  as a function of SNP-level annotation  $a_j$

$$\log_{10} \left( \frac{\pi_j}{1-\pi_j} \right) := \theta_0 + a_j \theta$$

## Data-adaptive threshold:

- Estimate enrichment parameter  $\theta$  from data
- “Enriched” data  $\rightsquigarrow \theta > 0 \rightsquigarrow$  larger  $\pi_j$  for  $a_j = 1 \rightsquigarrow$  increased power
- “Null” data  $\rightsquigarrow \theta \approx 0 \rightsquigarrow$  unchanged  $\pi_j \rightsquigarrow$  maintained type 1 error

**References:** Veyrieras *et al.* (2008); Carbonetto & Stephens (2013); Pickrell (2014); Kichaev *et al.* (2014); Chen *et al.* (2016); Li & Kellis (2016); Wen *et al.* (2017).



# Address Issue 2: Modeling linkage disequilibrium

**Genome-wide multiple-SNP likelihood function:**

$$L_{\text{RSS}}(\beta) := \text{Normal}(\hat{\beta}; \hat{S}\hat{R}\hat{S}^{-1}\beta, \hat{S}\hat{R}\hat{S})$$

- multiple-SNP parameter:  $\beta := (\beta_1, \dots, \beta_p)'$
- single-SNP summary data:  $\hat{\beta} := (\hat{\beta}_1, \dots, \hat{\beta}_p)'$
- $\hat{S} := \text{diag}(\hat{s}), \hat{s} := (\hat{s}_1, \dots, \hat{s}_p)', \hat{s}_j^2 := \hat{\sigma}_j^2 + n^{-1}\hat{\beta}_j^2 \simeq \hat{\sigma}_j^2$
- $\hat{R}$ : shrinkage estimate of LD (Wen & Stephens, 2010)
- **“Big data”** genetics: jointly analyze 1.1 million SNPs

**Reference:** Zhu and Stephens (2017 To appear). <http://dx.doi.org/10.1101/042457>.

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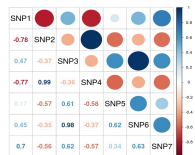
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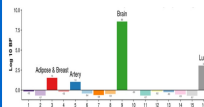
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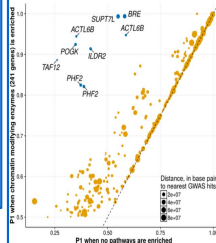
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$$\rho(\beta | \text{Data}, M_1) \text{ vs } \rho(\beta | \text{Data}, M_0)$$

# We apply the method to analyze 31 complex traits and 4,026 gene sets.

## This application is not small:

**# of Parameters =  $31 \times (3,913+113) \times 1.1 \text{ Million} \approx 137 \text{ Billion}$**

- **31** human phenotypes
- **3,913** biological pathways
- **113** tissue-based gene sets
- **1.1 million** common SNPs

## One student can get this done, aided by:

- Publicly available input data (GWAS & LD & gene sets)
- Variational inference; Squared extrapolation method (SQUAREM)
- Parallel computing; Hierarchical data format (HDF5)

# Our full results are publicly available.

## ■ **Results**

<https://xiangzhu.github.io/rss-gsea/results>

## ■ **Software**

<https://github.com/stephenslab/rss>

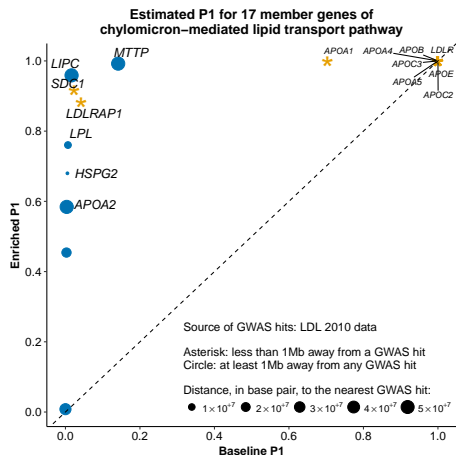
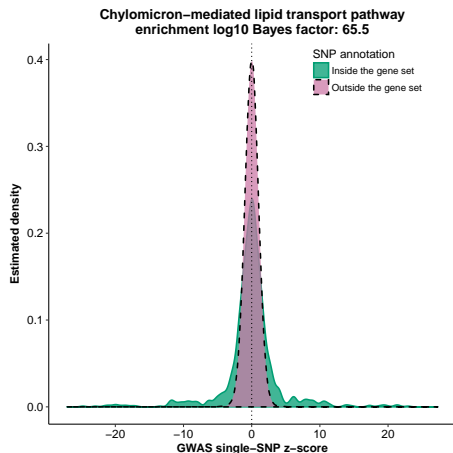
## ■ **Demonstration**

<https://stephenslab.github.io/rss/Example-5>

## ■ **R package (in progress)**

<https://github.com/stephenslab/rssr> (N. Knoblauch)

# Example: Low-density lipoprotein & *MTTP*



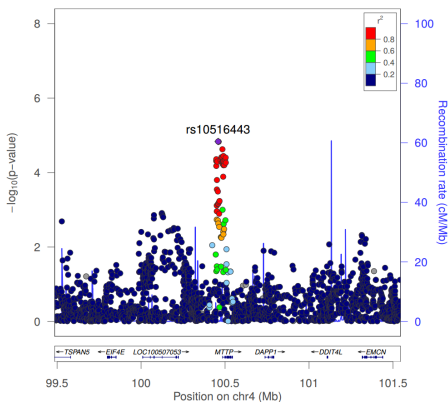
■  $P_1 := 1 - \text{Prob}(\beta_j = 0, \forall \text{ SNP } j \in \text{locus} \mid \text{Data})$

■ ***MTTP***: baseline  $P_1 = \mathbf{0.14}$  vs enriched  $P_1 = \mathbf{0.99}$

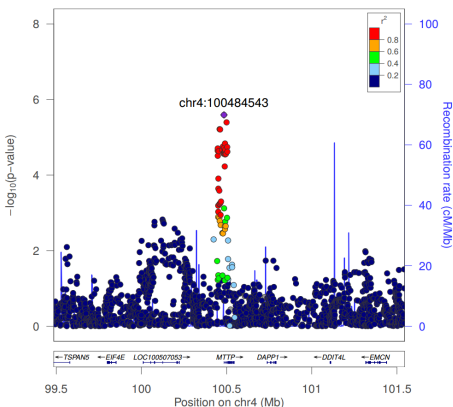
# Example: Low-density lipoprotein & *MTTP*

- Total sample size: **95,454** (2010) → **173,082** (2013)
- GWAS *p*-value:  **$1.5 \times 10^{-5}$**  (2010) →  **$2.6 \times 10^{-6}$**  (2013) “<”  $5 \times 10^{-8}$

LDL 2010 data, max sample size: 95,454



LDL 2013 data, max sample size: 173,082



**References:** Global Lipids Genetics Consortium (2013); Teslovich *et al.* (2010).

# Example: Alzheimer's disease & Liver

- Tissue-based gene sets
  - HE: highly expressed
  - SE: selectively expressed
  - DE: distinctively expressed

## ■ GTEx RNA-seq data

### ■ Top-enriched tissues

- Adrenal gland

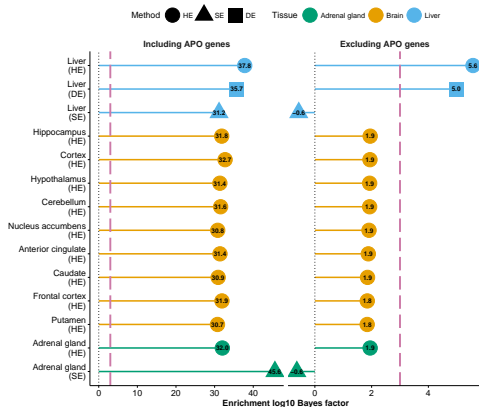
- Brain

- **Liver** (even w/o APO)

### ■ Non-APO, liver gene: **TTR**

- baseline  $P_1 = \mathbf{0.64}$

- enriched  $P_1 = \mathbf{1.00}$



**References:** Xi *et al.* (2017+); Dey *et al.* (2017);  
The GTEx Consortium (2015); Lambert *et al.* (2013).

# What's next?

## We develop a new enrichment analysis method that:

- uses **publicly available** data as input
- **efficiently** assesses thousands of gene sets
- **automatically** identifies trait-associated genes

## Future work:

- one annotation at a time → jointly analyze **many** annotations
- gene-based annotations → “**finer-scale**” annotations
- “spike-and-slab prior” → more **flexible** effect size distributions



# Acknowledgements

- Joint work with **Matthew Stephens**

- **GWAS Consortia**

Teslovich *et al.* (2010); Manning *et al.* (2012); Morris *et al.* (2012); van der Harst *et al.* (2012); Köttgen *et al.* (2013); Lambert *et al.* (2013); Okada *et al.* (2014); Ripke *et al.* (2014); Wood *et al.* (2014); Day *et al.* (2015); Liu *et al.* (2015); Locke *et al.* (2015); Nikpay *et al.* (2015); Shungin *et al.* (2015); Okbay *et al.* (2016); van Rheenen *et al.* (2016)

- **GTEx Consortium**

<https://gtexportal.org>

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**MOORE**  
FOUNDATION



THE UNIVERSITY OF CHICAGO  
**RESEARCH  
COMPUTING  
CENTER**

# Thank you!

- **Preprint:** <https://doi.org/10.1101/160770>
- **Software:** <https://github.com/stephenslab/rss>
- **Contact:** [xiangzhu\[at\]uchicago\[dot\]edu](mailto:xiangzhu[at]uchicago[dot]edu)